

Structures Solved Using MPEC Technologies (2005-present)

Target Name	Organism	Resolution Å	PDB ID	Data Method	R-Factor	Publication Date	Journal
Aquaporin Aqpm	<i>E. coli</i>	1.68	2F2B	X-ray	19.30%	2005	<i>PNAS</i>
Aquaporin Aqpm	<i>E. coli</i>	2.3	2EVU	X-ray	22.60%	2005	<i>PNAS</i>
Lac Permease (acidic)	<i>E. coli</i>	3.3	2CFP	X-ray	32.80%	2006	<i>EMBO</i>
Lac Permease (neutral)	<i>E. coli</i>	2.95	2CFQ	X-ray	29.80%	2006	<i>EMBO</i>
Lac Permease (w.type)	<i>E. coli</i>	3.6	2V8N	X-ray	33.30%	2007	<i>PNAS</i>
Glur2 Complex Cnqx	<i>R. norvegicus</i>	2.5	3B7D	X-ray	28.30%	2007	<i>Science</i>
Amtb Glnkx complex	<i>E. coli</i>	1.96	2NS1	X-ray	19.80%	2007	<i>PNAS</i>
Aqpz Mutant T183c	<i>E. coli</i>	2.3	2O9D	X-ray	23.80%	2007	<i>JMB</i>
AqpZ T183C mercury	<i>E. coli</i>	2.2	2O9E	X-ray	24.30%	2007	<i>JMB</i>
AqpZ mutant L170C	<i>E. coli</i>	2.55	2O9F	X-ray	28.60%	2007	<i>JMB</i>
AqpZ L170C Hg	<i>E. coli</i>	1.9	2O9G	X-ray	19.50%	2007	<i>JMB</i>
Aquaglyceroporin PfAQP)	<i>P. falciparum</i>	2.05	3C02	X-ray	30.60%	2008	<i>Nat. Struct. Mol. Bio.</i>
Rh-like protein	<i>N. europaea</i>	1.99	3BHS	X-ray	23.70%	2009	<i>PDB</i>
Aquaporin 4	<i>H. sapiens</i>	1.8	3GD8	X-ray	16.50%	2009	<i>PNAS</i>
RhCG	<i>H. sapiens</i>	2.1	3HD6	X-ray	19.50%	2010	<i>PNAS</i>
Sec 61 α translocon	<i>P. furiosus</i>	2.9	3MP7	X-ray	31.70%	2010	<i>PNAS</i>
AcrB pump +peptide	<i>E. coli</i>	3.3	refining	X-ray	-		
ZneB	<i>E. coli</i>	2.8	3LNN	X-ray	31.40%	2010	<i>PNAS</i>
Aqp from <i>A. fulgidus</i>	<i>A. fulgidus</i>	3.0	3NE2	X-ray	27.40%*		
CcmG thioredoxin	<i>E. coli</i>	2.3	3K8N	X-ray	25.54%*		
OmpF porin	<i>E. coli</i>	3.79	3K19	X-ray	28.80%	2010	<i>Protein Science</i>
OmpF porin	<i>E. coli</i>	4.39	3K1B	X-ray	32.90%	2010	<i>Protein Science</i>
Cytoplasmic domain HKR QseQ	<i>E. coli</i>	2.5	3JZ3	X-ray	26.20%	2010	<i>Protein Pept. Lett.</i>
N-terminal domain of GluN1	<i>R. norvegicus</i>	3.4	3Q41	X-ray	27.90%	2011	<i>J. Neuroscience</i>
Yeast erg11	<i>S.cerevisiae</i>	2.1		X-Ray		2011	<i>In preparation</i>
ZneA (+ Zn2+) form 1	<i>C.metallidurans</i>	3.0		X-ray	32.9%*	2011	<i>In preparation</i>
ZneA (+Zn2+) form 2	<i>C.metallidurans</i>	3.8		X-ray	33.7%*	2011	<i>In preparation</i>
Gramicidin	<i>B. subtilis</i>	1.7	2xdc	X-ray	18.10%	2010	<i>Biophys. J.</i>

Gramicidin	<i>B. subtilis</i>	1.08	2y5m	X-ray	13.20%	2011	<i>Cryst. Growth Des.</i>
Gramicidin	<i>B. subtilis</i>	1.26	2Y6N	X-ray	14.9%	2011	<i>Cryst. Growth Des.</i>
Cytochrome caa3 oxidase	<i>T. thermophilus</i>	2.35	2yev	X-ray		2011	<i>In preparation</i>
B2-adrenoreceptor	<i>Human</i>	3.5	3pds	X-ray	24.10%	2011	<i>Nature</i>
B2AR-Gs complex	<i>Human</i>	3.2	3sn6	X-ray	22.80%	2011	<i>Nature</i>

*denotes in refinement

MPEC List of Commercialized Instruments and Chemicals, and Protocols Established (2006-present)

TECHNOLOGY DEVELOPMENT/ACHIEVEMENT		FINDINGS/USAGE/COMMENTS
Protein Production		
1	Established inducible, selectable overexpressing human integral membrane proteins in transient, and stably transformed Human Embryonic Kidney 293S cells	Protocols for overexpression of human and eukaryotic membrane proteins in HEK (human) cells engineered to add only core sugars at glycosylation sites for homogeneity. Published.
2	Established efficient expression screening of human membrane proteins in transiently transfected Human Embryonic Kidney 293S cells.	Screening of proteins produced in HEK cells using fusion to fluorescent proteins to assay expression levels. Published.
3	Established a robust expression system in <i>S.cerevisiae</i> for membrane proteins.	Useful for expression of eukaryotic membrane proteins using a suite of vectors including those fused to cleavable e oligo-histidine tags, fluorescent proteins, yeast signal sequences, Precision protease sites.
4	Established baculovirus expression system	
5	Established standard protocol for expression, purification, and characterization of stable membrane protein preparation for structural studies.	
6	Established robust expression of human AQP4 for use in therapeutic applications against Neuromyelitis Optica.	Protein production that feed the community of researchers who study this rare but lethal autoimmune disease. Provision for the Guthy-Jackson Foundation (NMO).
7.	Established robot for screening crystallization plates by X-ray diffraction at the ALS	
8.	Established an E. coli-based cell-free expression system for integral membrane protein production and reconstitution into a bicontinuous lipidic mesophase	For use in tandem expression and crystallization of integral membrane proteins
Stabilization Technologies		
8.	Synthesis and characterization of new facial detergents	In collaboration with Dr S. Gellman produced dimeric facial amphiphiles. Published
9.	Novel lipids synthesized and purified at the gram scale for use in in meso crystallization trials	
10	Protocol: phage display panning to select Fragments antigen binding (Fabs) against membrane proteins	A naïve B cell library, expressed on phage, optimized for membrane proteins successfully used against various membrane proteins (published)
11	Protocol: ELISA based characterization of phage display selected Fabs	Allows rapid and reliable characterization of Fab binding, using ELISA and unpurified Fabs, and thus eliminates false positives or weaker binders before Fab production (published)
Biophysical Characterization		
12	Protocol for determining membrane protein subunit stoichiometry and homogeneity while also quantifying associated detergent.	Use of light scattering, refractive index plus viscometry and concentration measurements allows determination of the oligomeric state of an integral membrane protein in detergent solution. (Tetra detector)
13	Protocol - fluorescence assisted size exclusion chromatography (FSEC)	Allows for characterization of expression levels of membrane proteins in <i>S.cerevisiae</i> , HEK293 cells, and <i>E.coli</i>

14	Protocol - fluorescence assisted size exclusion chromatography (TD-FSEC)	Allows for characterization of expression levels of membrane proteins in E.coli,
15	Protocol for Mass Spectrometric Analysis of Membrane Proteins	Allows analysis of integral membrane proteins directly in the presence of lipid and detergent.
16	Protocol for assay of thermal stabilization in 96 well format using qPCR Viia 7 from ABI/Invitrogen.	Assay protocols adapted to measure effects on structure detected by melting curves and stabilization induced by lipids, detergents, buffers and ligands. Dyes co-developed to match qPCR detectors for MP stability measure.
17	Validation of Fab-membrane protein interaction by electron microscopy (EM)	In collaboration with Dr. Yifan Cheng, interactions between phage display selected Fabs and membrane proteins are validated by negative stain EM. Distinctive images of Fabs in negative staining allow easy detection of Fab-membrane protein complexes and thus visual confirmation of complex formation.
18	Established a host lipid screen for use in membrane protein crystallization in lipidic mesophases	Used to extend the range of membrane protein types that yield diffraction quality crystals by the in meso crystallization method
19	Established a high-throughput protocol for assaying the enzymatic activity of integral membrane proteins reconstituted into the lipidic cubic mesophase	Useful in evaluating functional activity of membrane proteins in meso as a prelude to crystallization
20	Established X-ray screening in crystallization plates	Robot designed in operation at ALS synchrotron Berkeley.
21	Automated crystal handling tasks at the ALS	http://bl831.als.lbl.gov/~jamesh/beamline/holton_utilities.html
22	Program that informs on feasibility of structure determination	http://bl831.als.lbl.gov/xtalsize.html
Technology Integration		
23	Pipeline for production in <i>S.cerevisia</i> MPEC vector suite through solubilization, purification, characterization, crystallization and structure determination.	Integration of all protocols/technologies into a pipeline for the efficient determination of the structure of membrane proteins.
Technology Validation		
24	Validation of technology	Structure solution of over 25 membrane proteins including mutant and variant species using MPEC technologies.

Outreach and Training (2006-Present)

Name	Affiliation	Dates	Purpose/Training
Chantal Eckende	Univ Brussels	May 2008 –Jan 2010 Spent at UCSF	Expression Purification ZneA/B/C
Guthy-Jackson Foundation for NMO disease	Private Not-for Profit Research Foundation	2009-present	Contracted to supply 150mg/year pure human AQP4, target of the lethal orphan disease Neuromyelitis Optica
Andre Bachmann	University of hawaii	2008	Human 4 crossing Golgi trafficking protein, now in crystal trials
Niek Dekker	Astra Zeneca Sweden	Provided MP expression systems and purification	Ion Channels, GPCRs, TRP channels Expression
Brian Monk	University of Auckland NZ	Jan 2010-Nov 2010 Spent at UCSF	Yeast expression Purify, Crystallize yeast ABC transporters, and cyt oxidases erg11: crystal structure determined
Richard Cannon	University of Auckland NZ	Feb 2009- Oct 2009 Spent at UCSF	Yeast expression Purify fungal ABC drug exporters.
Erwin Lamping		Feb 2009-Sep 2009 Spent at UCSF	Yeast expression Purify fungal ABC drug exporters.
Cedric Govaerts	Universite libre de Bruxelles (belgium)	2009-	LMRP a multi-drug resistant transporter protein
Nicole Tischler	Fundacion Ciencia para la vida Chile	2009-	Andes virus envelope glycoprotein Gc Preparation
Ignaccio Munoz	Fundacion Ciencia para la vida Chile	May 2009- Sept 2010 Spent at UCSF	Andes virus envelope glycoprotein Gc Preparation
Claudia Mergel	Goethe University in Frankfurt	Mar 2010-Nov 2010 Spent at UCSF	Protein purification, crystallization, succeeded
Sebastian Stiller	Goethe University in Frankfurt	June2010-Sep 2010 Spent at UCSF	Protein purification, crystallization, succeeded
Mickie Cheng	UCSF	July 2011-present	Membrane protein purification training
Natalia Jura	UCSF	Dec 2011-present	Membrane protein purification training
Daniel Süveges	UCSF	Dec 2011-present	Membrane protein purification training
Teresa Garrett	Vassar College	Spent at UCSF	
Mickie Cheng	UCSF	July 2011-present	Membrane protein purification training
Natalia Jura	UCSF	Dec 2011-present	Membrane protein purification training
Daniel Süveges	UCSF	Dec 2011-present	Membrane protein purification training
20 students	GPCR workshop, Maui	Dec 2011	In meso crystallization training /MC
20 students	ACMC11, Melbourne, Australia	Dec 2011	In meso crystallization training /MC
20 Students	University of Helsinki, Finland	May 2011	In meso crystallization training /MC
20 Students	University of Granada, Spain	May 2011	In meso crystallization training /MC
V Srinivasan	Free University of Brussels	April 2011	In meso crystallization training /MC

H Bridges	University of Cambridge	April 2011	In meso crystallization training /MC
F Jaenecke	Halle-Wuttenburg University	January 2011	In meso crystallization training

External Collaborations (outside MPEC)

Collaborator	Affiliation	Description of study	Target
Krieger, Monty	MIT	Purification & Structure solution	HDL Receptor –now crystallized
Bachmann, Andre	University of Hawaii	Protein Production Structure determination	Human 4 crossing Golgi trafficking protein, now in crystal trials; pediatric cancer target
Belogradov, Gregori	UCLA	Structure determination	Mitochondrial factor B; structure determined. Paper in <i>PNAS</i> 2008
Nicoll, Roger	UCSF, Cellular & Molecular Pharm.	Structure determination of AMPA receptor drug interactions.	Four membrane proteins in the AMPA receptor protein family and accessory TARPs; Paper in <i>Science</i> 2007
Farese, Robert	UCSF, Gladstone Institute	Towards Structure determination	Yeast protein DGA1 (Diacylglycerol acyltransferase)
Weissman, Jonathan	UCSF, Cellular & Molecular Pharm	Towards Structure determination: Now joined MPEC in 2009	Long chain fatty acid synthetase, 4 yeast proteins, in the FEN, SUR class. Now in crystallization trials
Detitta, George	Hauptmann Woodward Institute, Buffalo, NY	Our Membrane Proteins, screening in HT mode in Buffalo	Automated, high throughput microbatch under oil crystallization
Ismagilov, Rustem	University of Chicago	Protein supplied to develop Microcapillary crystallization	High throughput capillary crystallization
Lebon, Florence	UC Belgium	Protein Production Structure determination	Epilepsy target for drug discovery SV class
Giacomini, Kathleen	Pharmacogenomics Center, UCSF	Human Transporters to Structures	28 plasma membrane human transporters
Derisi, Joe	UCSF, Biochemistry Dept	Protein Production Structure determination	Chloroquin transporter and other MPs from <i>P.falciparum</i>
Rosenzweig, Amy	Northwestern Univ.	Protein Production Structure determination	Human metal trafficking proteins
Kossiakoff, Tony	University of Chicago	Manufacture of single chain, phage optimized antibodies for crystallization;	TRP channels
Rout, Michael	The Rockefeller University	Femtosecond structure determination	Expression and crystallization of nuclear pore complexes proteins
Kenis, Paul	University of Illinois Urbana-Champaign	Technology transfer and testing of his method for membrane proteins	Microfluidic crystallization
Gellman, Samuel	University of Wisconsin	Structure solution of facial amphiphiles	Tripod detergents, published
Dekker, Niek	Astra Zeneca, Sweden	Structure solution; Yeast Expression Systems	Ion Channels, GPCRs, TRP channels
Govaerts, Cedric F	Universite Libre de Bruxelles (belgium)	Structure solution	LMRP a multi-drug resistant transporter protein
Tischler, Nicole	Fundacion Ciencia Para la Vida, Chile	Structure solution	Andes virus envelope glycoprotein Gc
Yu, Edward	University of Southern California, School of Medicine	Protein Production Structure determination	Protein production of human Claudins
Kossiakoff, Tony	University of Chicago	Protocols for anti-membrane protein antibodies	TRP channels
VandenBussche, Guy	Universite do Bruxelles Belgium	July 2007-ongoing	Pipeline and methods transfer
Cannon, Richard	University of Auckland NZ	Sabbatical leave 6 mos at MPEC. Protein production purification crystallization.	Transporters in <i>S.cerevisiae</i>
Monk, Brian	University of Auckland NZ	Sabbatical leave 9 mos at MPEC. Protein production purification crystallization and structure.	Erg 11, drug complexes structures

Tampe, Robert	Goethe University in Frankfurt	Sabbatical leave 9 mos at MPEC. Protein production purification crystallization.	T.t ABC transporter
Fromme, Petra	University of Arizona	Crystals for Free Electron Laser Diffraction	
Parsons, Stanley	UCSB	Protein Production Structure determination	Vesicular Ach Transporter
Cassida, John	UCB	Expression and Structure determination	GABA activated channels
Koth, Christopher	Genentech	X-ray crystallography	Class B GPCR
Kurtz, Ira	UCLA	X-ray crystallography	AE1 (Band 3)
Immke, David	Amgen	X-ray crystallography	Class B GPCRs and phosphate transporter
Vandenbussche, Guy	Université Libre de Bruxelles	X-ray crystallography	Heavy metal transporters
Ekende, Chantal	Université Libre de Bruxelles	X-ray crystallography	Heavy metal transporters
Sali, Andrej	UCSF	In silico drug screening	Glucagon Receptor
Bonomi, Massimiliano	UCSF	In silico drug screening	Glucagon Receptor
Zur, Arik	UCSF	In vivo drug screening	Glucagon Receptor
Raetz, Chris	Duke University	Lipid A biosynthetic enzymes	Glycosyl transferase & hydrolase
Sill, Anita	UCSF	Fungal pathogenesis	Fungal pathogen factor
Lee, John K.	University of Minnesota	Purification of human membrane protein targets	Claudins
Hays, Frank	University of Oklahoma	Assay and purification of membrane proteins in yeast.	Nucleoside and sulfate transporter
Garrett, Teresa	Vassar College	Assay of target	
Cheng, Yifan	UCSF	Validation of Fab-membrane protein interactions by EM	Glutamate transporter homologue and ABC transporter

Publications (2005 – present)

2011 (32 publications)

83. Kim, J., Stroud, R.M. and Craik, C.S. Rapid identification of recombinant Fabs that bind to membrane proteins. *Methods* Sep 20, 2011 *in press*
82. Schneider, E., Lee, M.S., Baharuddin, A., Goetz, D., Farady, C.J., Ward, M., Wang, C., Craik, C.S. A Reverse Binding Motif that Contributes to Specific Protease Inhibition by Antibodies. *J. Molecular Biology* Nov. 26, 2011 *in press*
81. Chaudhary, S.*, Pak, J.E.*, Gruswitz, F., Sharma, V., and Stroud, R.M. Overexpressing human integral membrane proteins in stably transformed Human Embryonic Kidney 293S cells. *Nature Protocols*, *in press* (2012). (*denotes equal contribution)
80. Chaudhary, S.*, Pak, J.E.*, Pedersen, B.P., Bang, L.J., Zhang, L.B., Ngaw, S.M.M., Green, R.G., Sharma V., and Stroud, R.M. Efficient expression screening of human membrane proteins in transiently transfected Human Embryonic Kidney 293S cells. *Methods*, *in press* (2012). (*denotes equal contribution)
79. Yang, D., Cwynar, V.A., Hart, D.J., Madanmohan, J., Lee, J., Lyons, J., Caffrey, M. Preparation of 1-Monoacylglycerols via the Suzuki-Miyaura Reaction: 2,3-Dihydroxypropyl (Z)-tetradec-7-enoate. *Org. Synth.* 89: *in press* (2012)
78. Wu, S.*, Avila-Sakar, A.*, Kim, J.*, Booth, D., Greenberg, C., Rossi, A., Liao, M., Alian, A., Griner, S., Juge, N., Mergel, C., Chaparro-Riggers, J., Strap, P., Tampé, R., Edwards, R.H., Stroud, R.M., Craik, C.S. and Cheng, Y. (2011). Fabs enable single particle cryoEM studies of small proteins *Structure submitted*. (*denotes equal contribution)
77. Welch, M., Villalobos, A., Gustafsson, C., Minshull, J. (2011). Designing Genes for Successful Protein Expression. *Methods Enzymology*. **498**, 43-66. (PMID: 21601673)
76. Chaudhary, S., Pak, J.E., Gruswitz, F., Sharma, V., Stroud, R.M. (2011). Overexpressing Human Membrane Proteins in Stably Transfected and Clonal Human Embryonic Kidney 293S Cells. *Nature Protocols*
75. Stroud, R.M. (2011). New tools in membrane protein determination. *F1000 Biol Rep.* 3:8. (PMID: 21655333 / PMCID: 3100781)
74. Stroud, R.M, Schertler, G.F. (2011). Membranes. *Curr Opin Struct Biol.* **21**, 495-6. (PMID: 21875531)
73. Stepanov, S., Hilgart, M., Yoder, D. W., Makarov, O., Becker, M., Sanishvili, R., Ogata, C. M., Venugopalan, N., Aragão, D., Caffrey, M., Smith, J. L., Fischetti, R. F. (2011). Fast fluorescence techniques for crystallography beamlines. *J. Appl. Cryst.* 44, 772-778. [doi:10.1107/S0021889811016748] (PMID: 21808424 / PMCID: PMC3140417) (Available at URL: <http://journals.iucr.org/j/issues/2011/04/00/dz5227/dz5227.pdf>)
72. Spence, J. C. H., Kirian, R. A., Wang, X., Weierstall, U., Schmidt, K. E., White, T., Barty, A., Chapman, H. N., Marchesini, S. & Holton, J. (2011). Phasing of coherent femtosecond X-ray diffraction from size-varying nanocrystals. *Opt. Express* 19, 2866-2873. (PMID: 21369108)
71. Smirnova, I., Kasho, V., Sugihara, J., Kaback, H.R. (2011). Opening the periplasmic cavity in lactose permease is the limiting step for sugar binding. *PNAS.* **108**, 15147-15151. (PMID: 21896727 / PMCID: PMC3174601)
70. Smirnova, I., Kasho, V., Kaback, H.R. (2011). LacY and the alternating access mechanism. *Biochemistry.* **45**, 9684-93 (PMID: 2199533 / PMCID: PMC3140417)
69. Shaya, D., Kreir, M., Robbins, R.A., Wong, S., Hammon, J., Bruggemann, A., and Minor, D.L. Jr. (2011). Voltage-gated sodium channel (NaV) protein dissection creates a set of functional ‘pore-only’ proteins. *Proc Natl Acad Sci. USA.* **108**, 12313-8. (PMCID: PMC3145705)
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- DeVree, B.T., et al. (2011). Structure and function of an irreversible agonist- β_2 adrenoceptor complex. *Nature*. **469**, 236-240. (PMCID: [PMC3074335](#))
67. Rasmussen, S.G.F., DeVree, B.T., Zou, Y., Kruse, A.C., Chung, K.Y., Kobilka, T.S., Thian, F.S., Chae, P.S., Pardon, E., Calinski, D., Mathiesen, J.M., Shah, S.T.A., Lyons, J.A., Caffrey, M., Gellman, S.H., Steyaert, J., Skiniotis, G., Weis, W.I., Sunahara, R.K., Kobilka, B.K. (2011). Crystal structure of the β_2 adrenergic receptor-Gs protein complex. *Nature*. **477**, 549-555. (PMCID: [PMC3184188](#))
66. Pye, V. E., Aragao, D., Lyons, J. A., Caffrey, M. (2011). Overview of International Conference on the Crystallization of Biological Macromolecules. *Cryst. Growth Des.* In press. (URL available when published) DOI: 10.1021/cg101379p
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64. Li, D., Lee, J., Caffrey, M. (2011). Crystallizing Membrane Proteins in Lipidic Mesophases. A Host Lipid Screen. *Cryst. Growth Des.* **11**, 530-537. (PMCID: [PMC3131202](#))
63. Kirian, R. A., White, T. A., Holton, J. M., Chapman, H. N., Fromme, P., Barty, A., Lomb, L., Aquila, A., Maia, F. R. N. C., Martin, A. V., Fromme, R., Wang, X., Hunter, M. S., Schmidt, K. E. & Spence, J. C. H. (2011). Structure-factor analysis of femtosecond microdiffraction patterns from protein nanocrystals. *Acta Cryst. A* **67**, 131-140. (PMCID: [PMC3066792](#))
62. Kim, J., Stroud, R.M. and Craik, C.S. (2011). Rapid identification of recombinant Fabs that bind to membrane proteins. *Methods*. (PMID: 21958987)
61. Kaback, H.R., Smirnova, I., Kasho, V., Nie, Y., Zhou, Y. (2011). The alternating access transport mechanism in LacY. *J Membr Biol*. **239**, 85-93. (PMCID: [PMC3030946](#))
60. Jiang, X., Nie, Y., Kaback, H.R. (2011). Site-directed alkylation studies with LacY provide evidence for the alternating access model of transport. *Biochemistry*, **50**, 1634-40. (PMID: [21254783](#) / PMCID: [PMC3057939](#) [Available on 2012/3/15])
59. Hofer, N., Aragao, D., J.A., Lyons, Caffrey, M. (2011). Membrane Protein Crystallization in Lipidic Mesophases. Hosting lipid affects on the crystallization and structure of a transmembrane peptide. *Cryst. Growth Des.* **11**, 1182-1192. (Available at URL <http://pubs.acs.org/doi/abs/10.1021/cg101384p>)
58. Hofer, N., Aragao, D., Caffrey, M. (2011). The Lipidic Cubic Phase as a Membrane Mimetic. *Biophys. J.* **100**, 2075.
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